

# Cocultivation of Algae and Bacteria for Improved Productivity and Metabolic Versatility

***Alex S. Beliaev***

***Pacific Northwest National Laboratory***

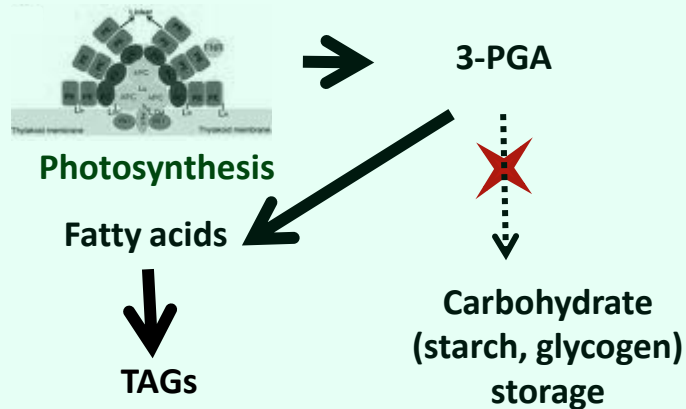
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# Axenic Cultures in Algal Biotechnology

## A. *Gene/pathway inactivation*



- Current approaches use axenic (pure) cultures of microalgae and/or cyanobacteria

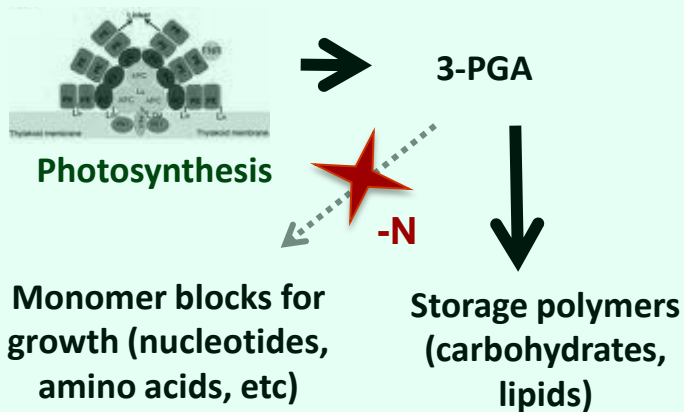
- Productivity is manipulated by imposing environmental or genetic perturbations

- *Examples:*

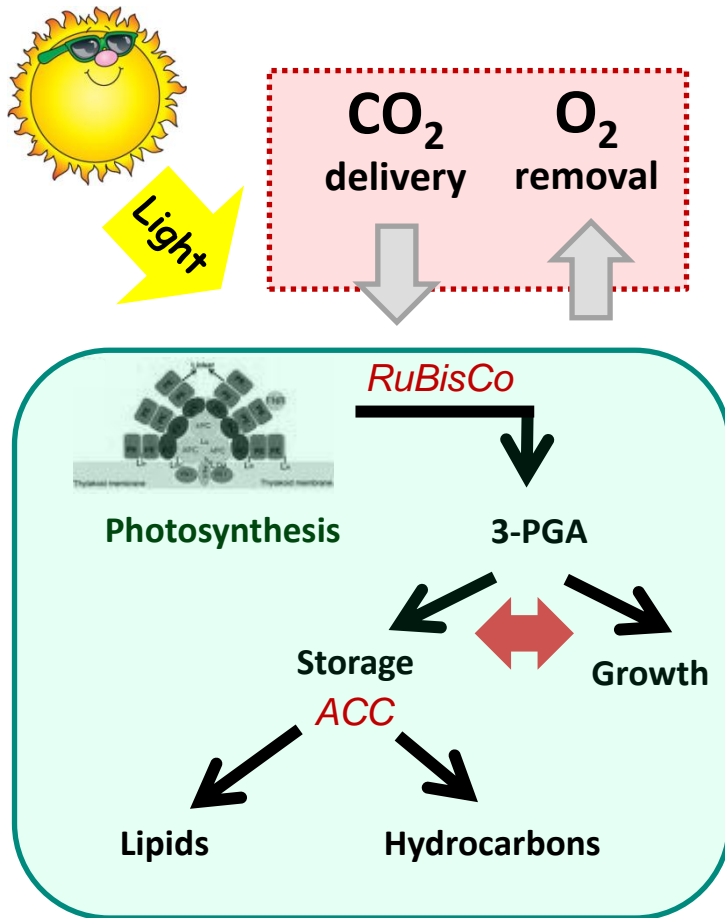
*A) Inactivation of competing pathways to redirect flux towards specific products*

*B) Nutrient (N, S) limitation to inhibit growth and enhance storage product accumulation*

## B. *Nutrient limitation*

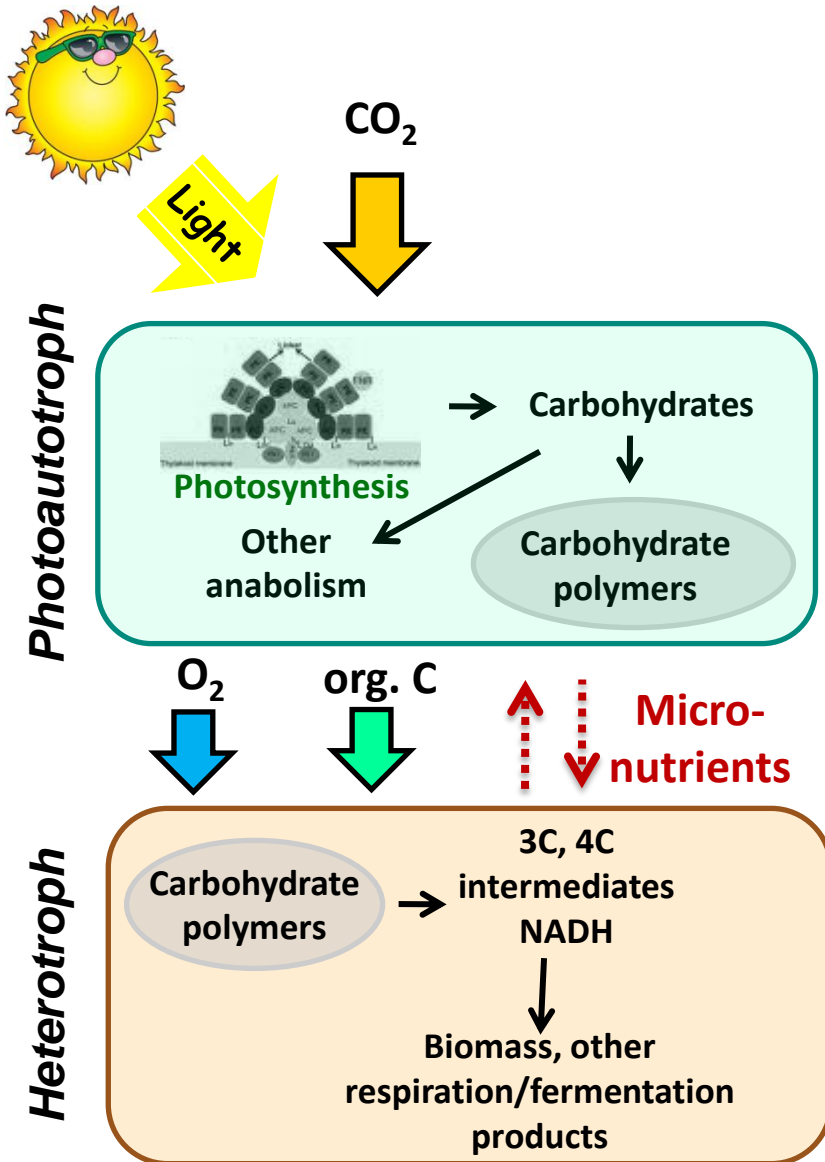


# Axenic Culture Challenges



- **Process engineering:** mass-transfer limitations involving gaseous substrate delivery (CO<sub>2</sub>) and product removal (O<sub>2</sub>)
- **Growth physiology:** balance the energy input with the downstream biosynthetic processes (growth vs. storage compounds)
- **Metabolic engineering:** coordination of various pathways needed; changes in expression and/or activity levels may have unanticipated secondary consequences upon product yields. Some functions are subject to product inhibition or allosteric regulation (*e.g.*, RuBisCo photorespiration; acetyl-CoA carboxylase regulation by palmitoyl-CoA).

# Co-Existence of Algae & Bacteria in Nature



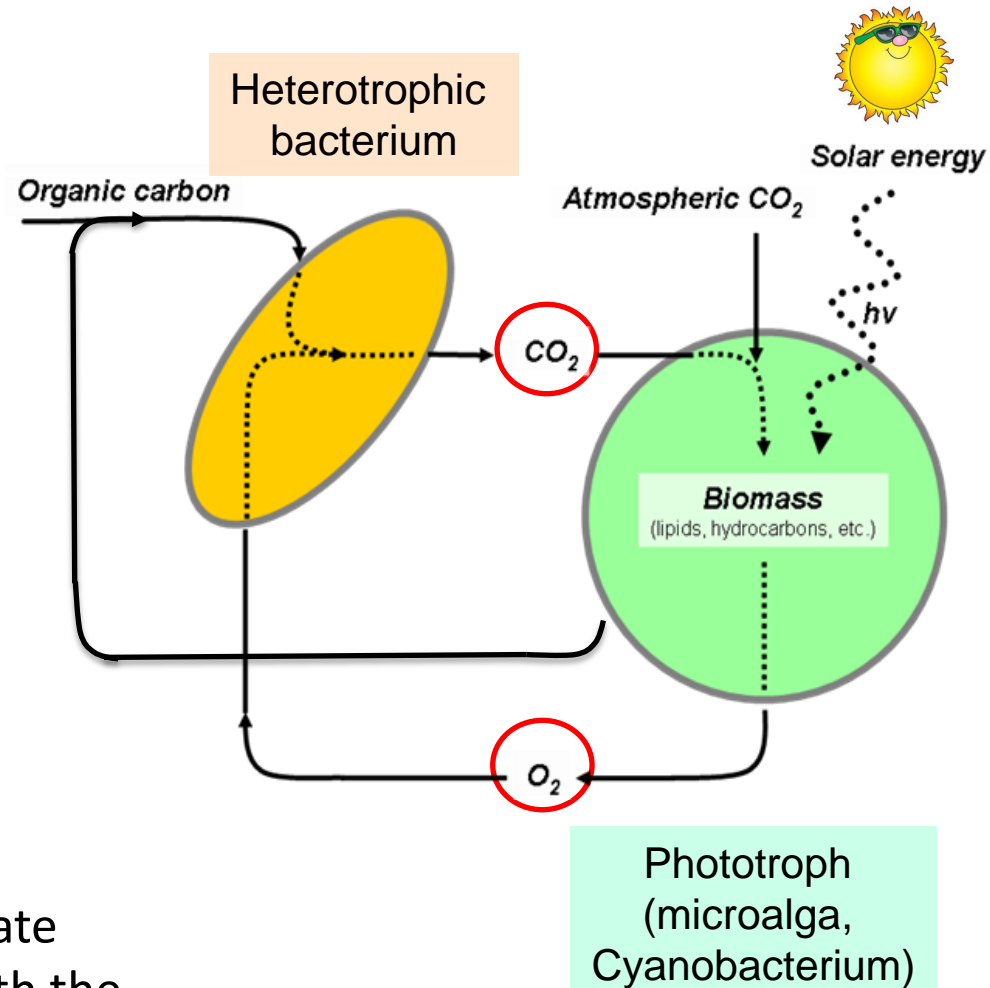
- Algae and cyanobacteria use sunlight and CO<sub>2</sub> and produce O<sub>2</sub> and C<sub>org</sub> molecules that support growth of heterotrophic bacteria
- Heterotrophic bacteria provide intrinsic stability and support growth of phototrophs by removing excess O<sub>2</sub>, increasing micro-nutrient availability, vitamin biosynthesis
- Algae-bacterial associations represent metabolically interactive, self-sustaining communities, which display adaptation to a range of harsh conditions

# Phototroph-Heterotroph Co-Cultures

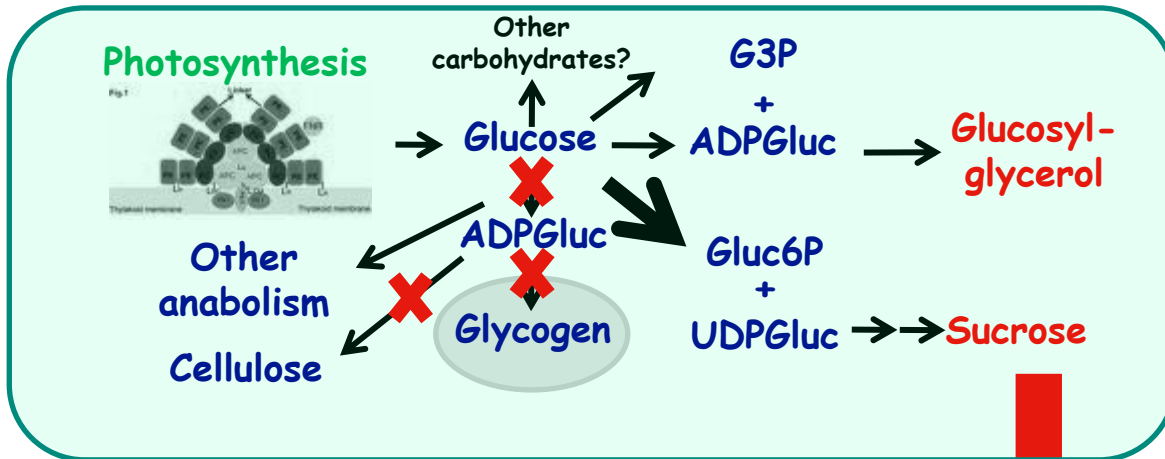
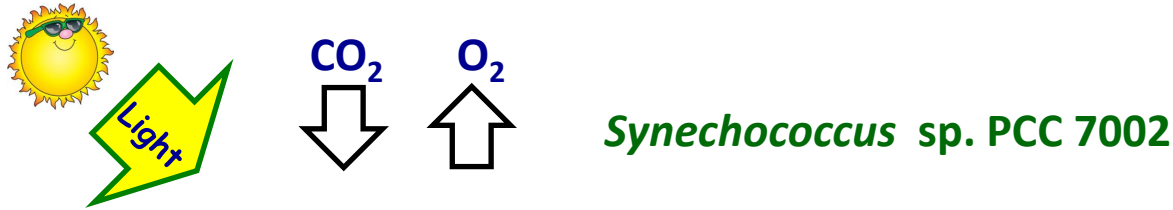
- **Metabolic coupling:**  $O_2$  produced by the algae is consumed by the heterotroph making stoichiometric amount of  $CO_2$  through oxidation of (endogenous or exogenous) organic C. Stoichiometric constraints drastically increase the intrinsic stability.

- **Advantages:**

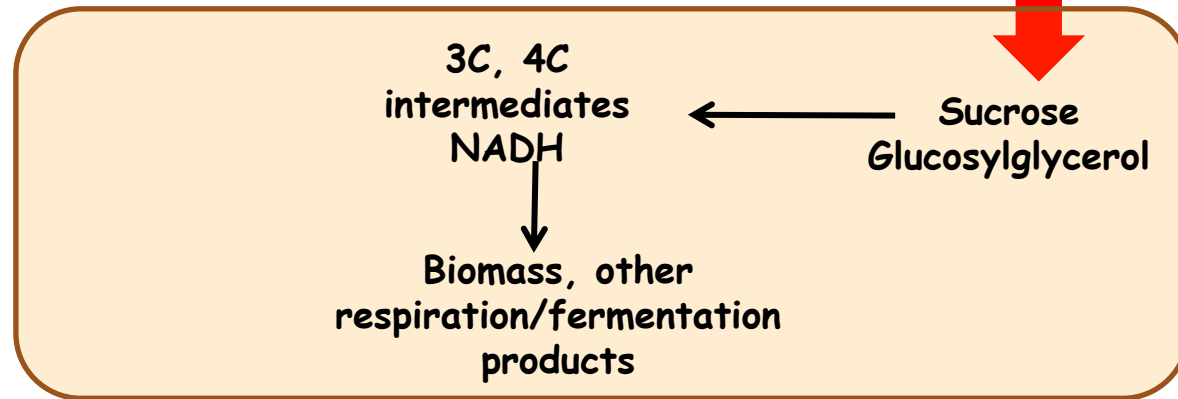
- Improved mass transfer & productivity
- Increased range of carbon sources
- Modularity & ability to spatially separate the processes of light &  $CO_2$  capture with the downstream photosynthate conversion



# Coupling through Photosynthate Secretion

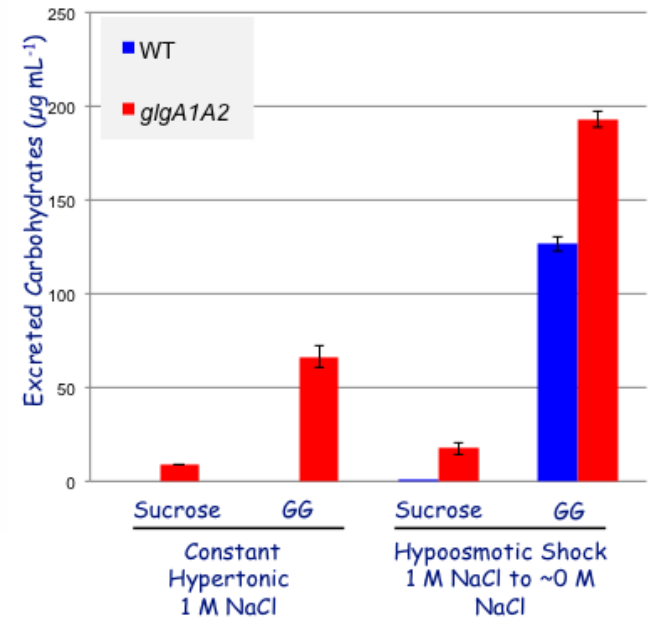
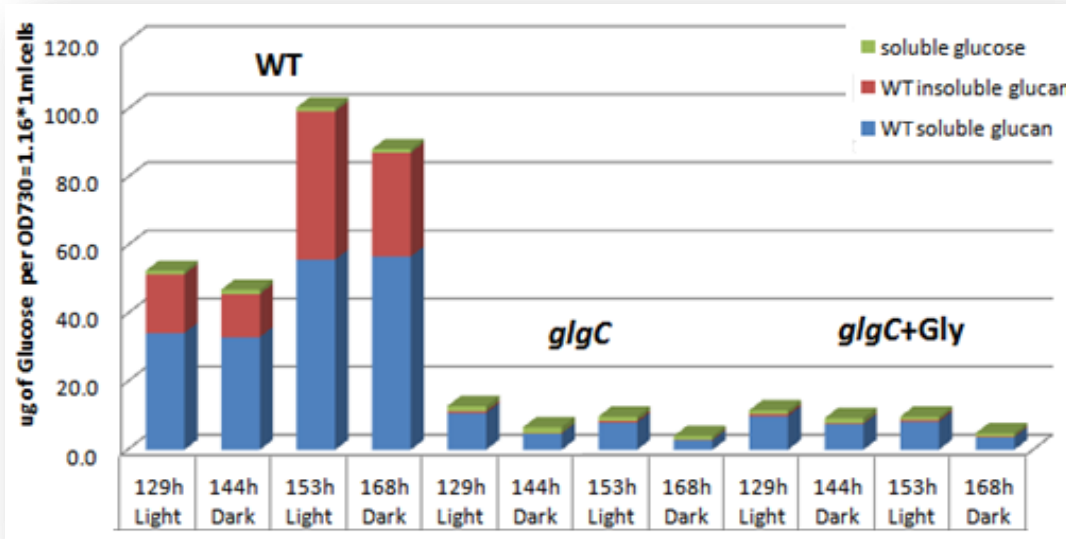


**Rationale:** Redirect fixed CO<sub>2</sub> to mono/ disaccharide derivatives, which can be excreted and used as a carbon and energy source for biofuel synthesis by heterotrophic organisms.



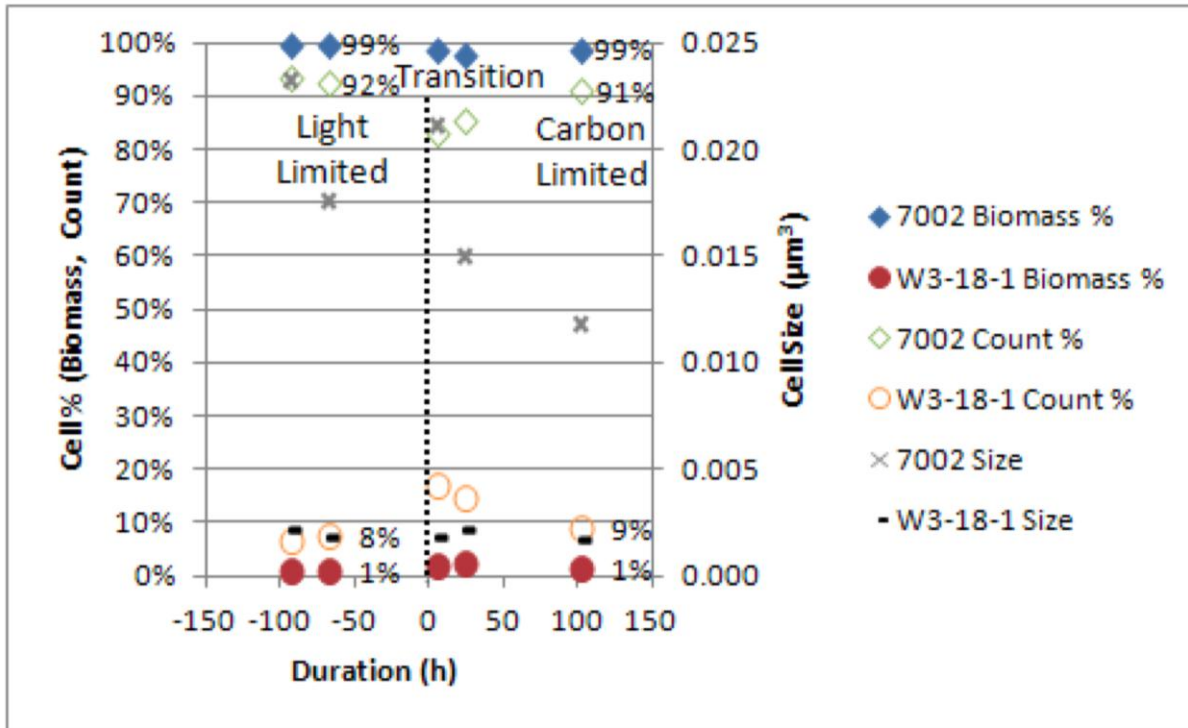
**Approach:** Eliminate glycogen storage by mutation of *glgA1*, *glgA2*, and *glgB*, and/or *glgC* but maintain high photosynthetic rate.

# Engineering Glycogen Metabolism to Increase Carbohydrate Excretion



This strategy works! Glucose, sucrose and glucosylglycerol are excreted in *glg* mutants of *Synechococcus* sp. PCC 7002 that cannot make glycogen.

# Metabolic Coupling through Secreted C



- Heterotrophic growth supported through secretion of sugars and osmolytes (>300hs)
- Biomass concentration can be manipulated by varying growth conditions (light, CO<sub>2</sub>)

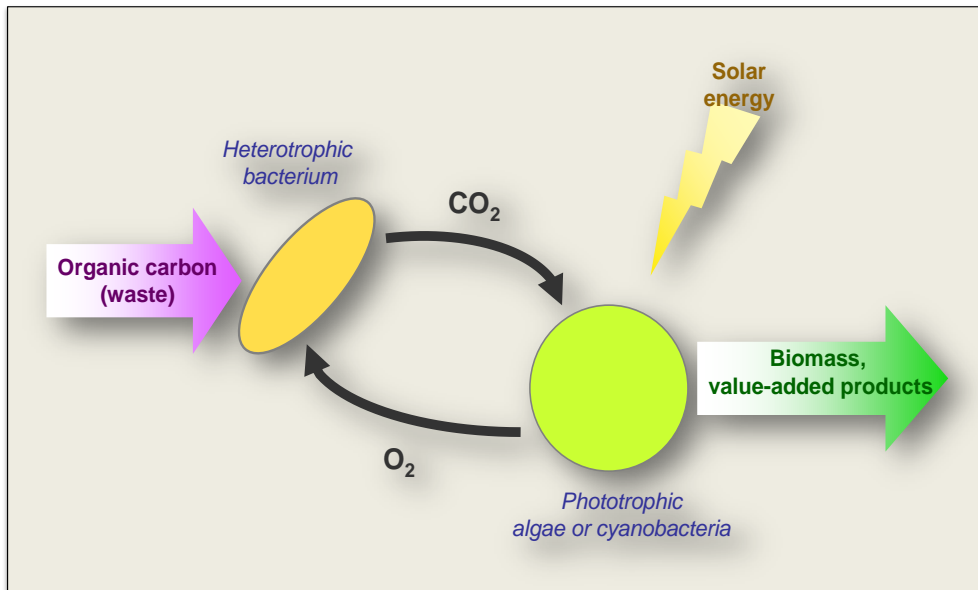
- Plug-and-play approach in which process of photosynthetic carbon fixation and product biosynthesis is spatially separated

Module A: CO<sub>2</sub> -> C<sub>org</sub> (sugars, organic acids)

Module B: C<sub>org</sub> -> target bio-product



# Metabolic Coupling via Exogenous C



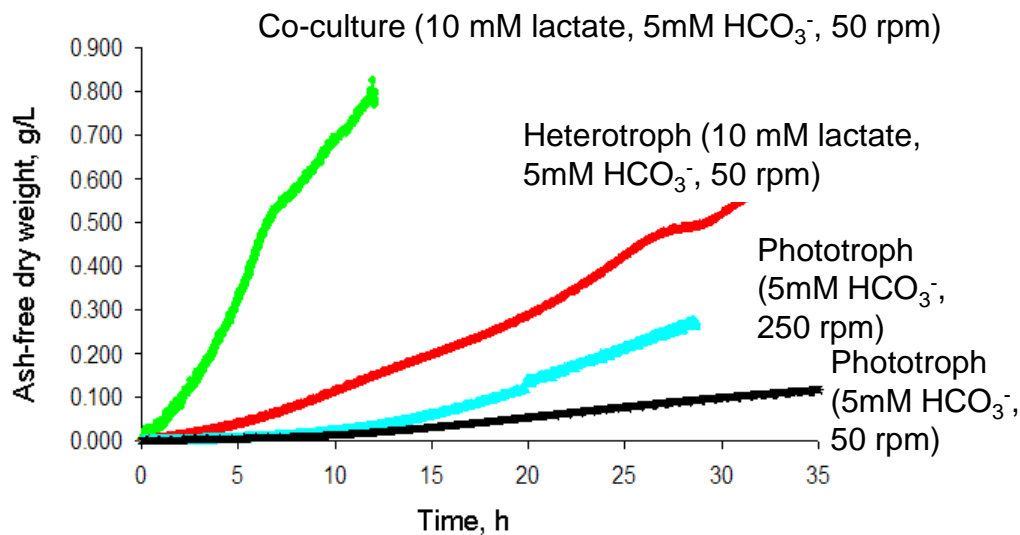
- Allows utilization of various C sources (including waste streams)

- Limited mass transfer as  $\text{O}_2$  and  $\text{CO}_2$  are produced throughout cultivation vessel

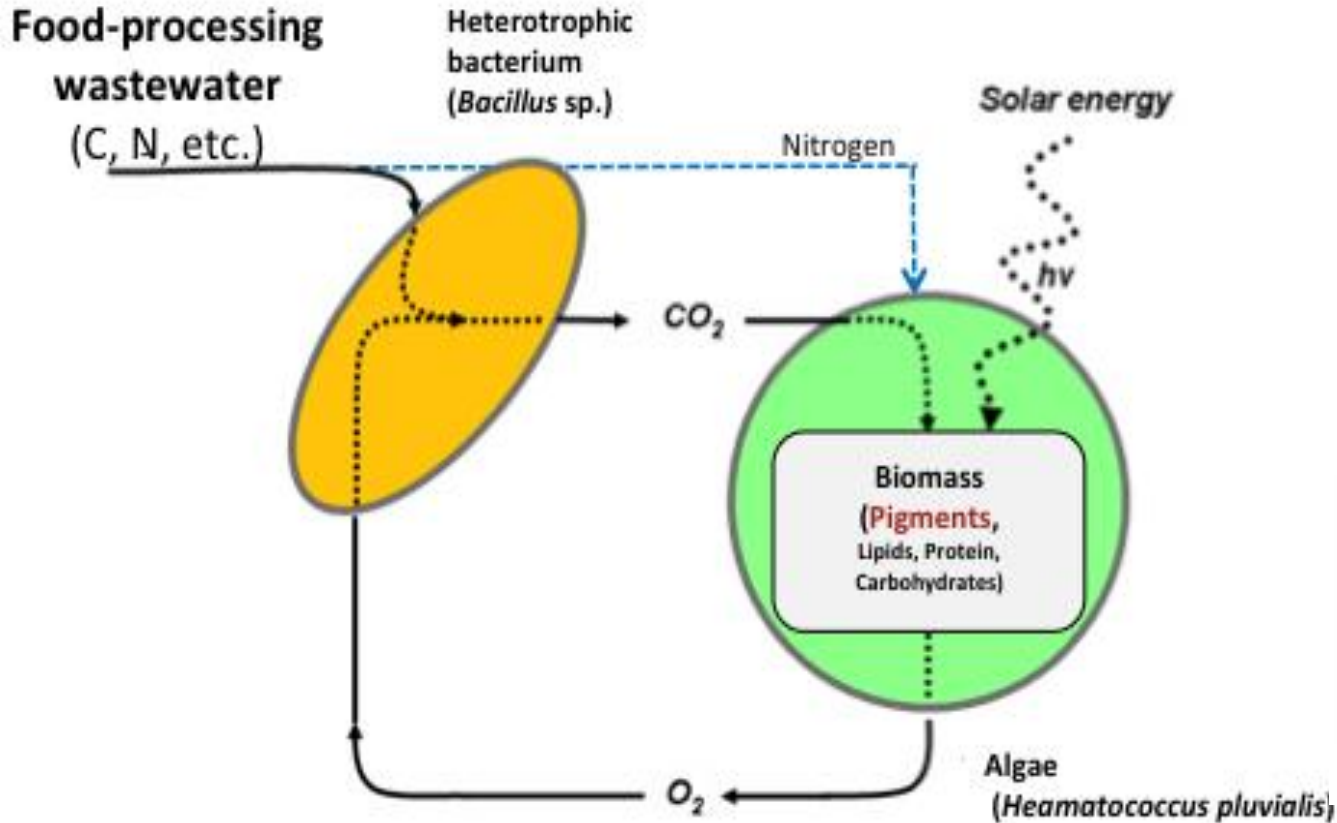
- Axenic (pure) cultures display significantly lower biomass productivity and growth rates

- Co-culture displays higher growth & productivity; does not need high mass transfer rates ; utilizes both carbon sources; no  $\text{O}_2$  accumulation

- Ratio of  $\text{C}_{\text{org}}/\text{CO}_2$  affects the proportion heterotroph & phototroph biomass

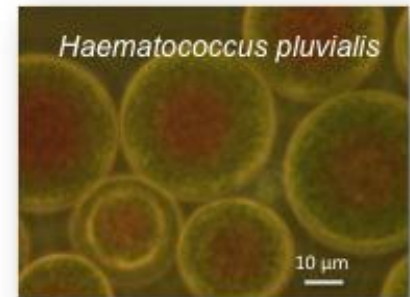


# Waste Treatment using Algal Co-cultures



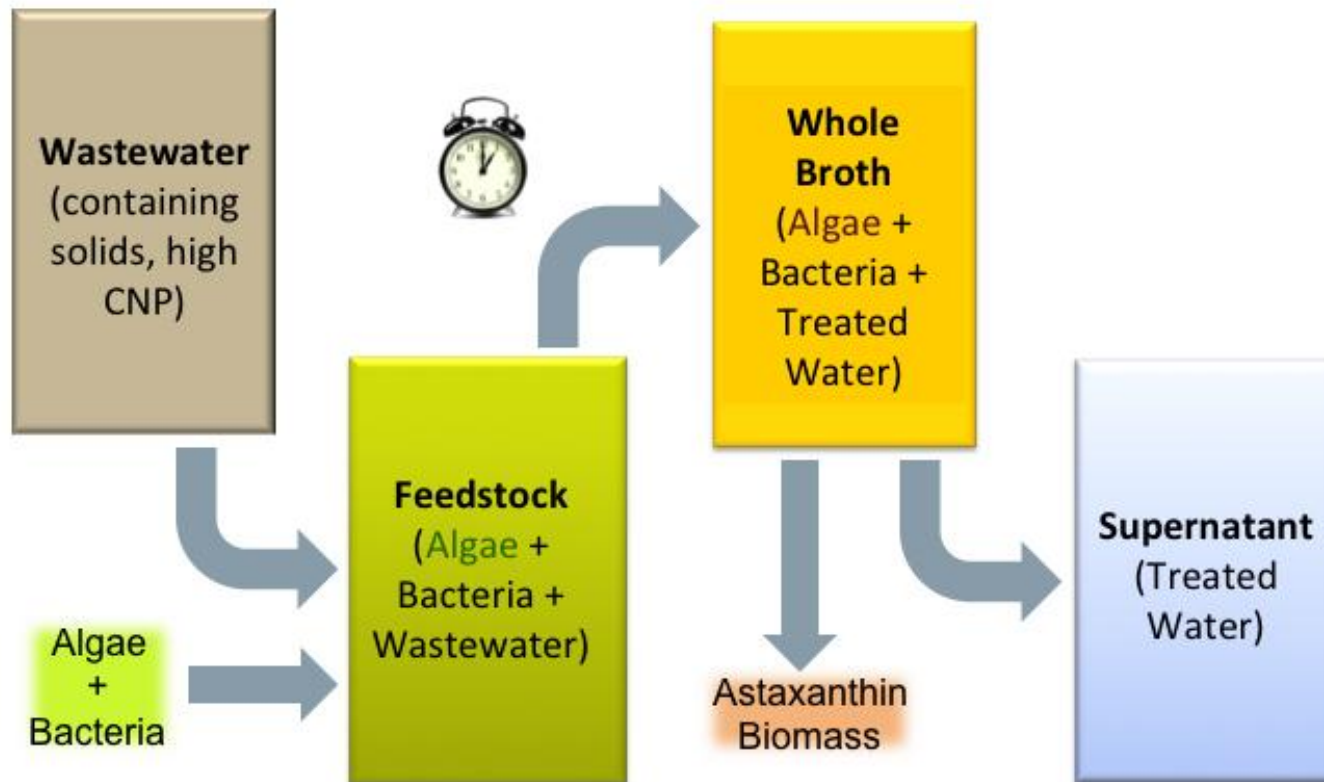
## Astaxanthin

- High-Value Pigment (~\$2000/kg)
- Feed: Salmon, Poultry
- Natural vs. Synthetic (95%)



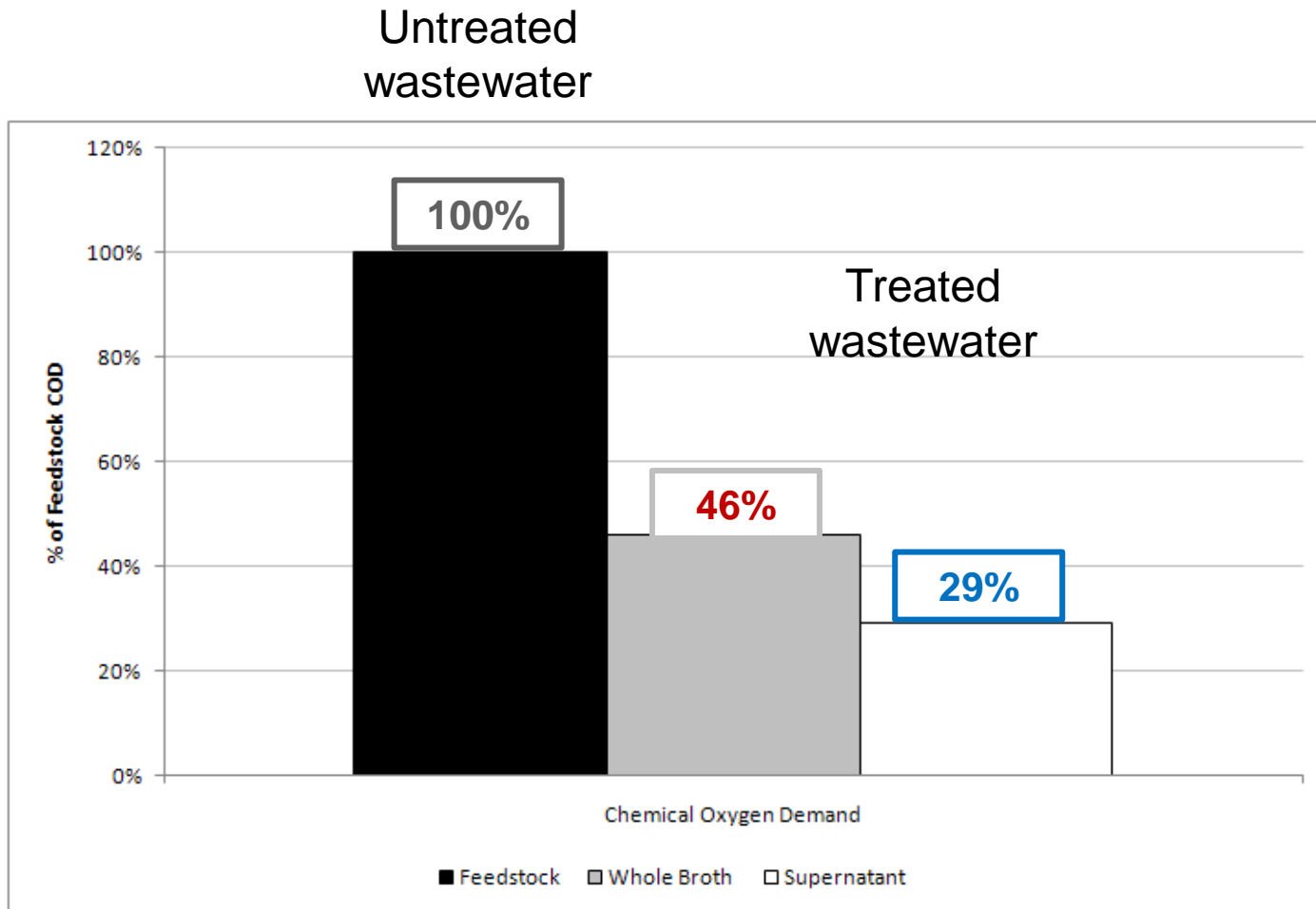
# Wastewater Treatment: Setup

- Wastewater with high concentration of complex carbohydrates, N, and P
- Co-culture *Bacillus* sp. and *Haematococcus pluvialis*
- Light, no bubbling, low agitation



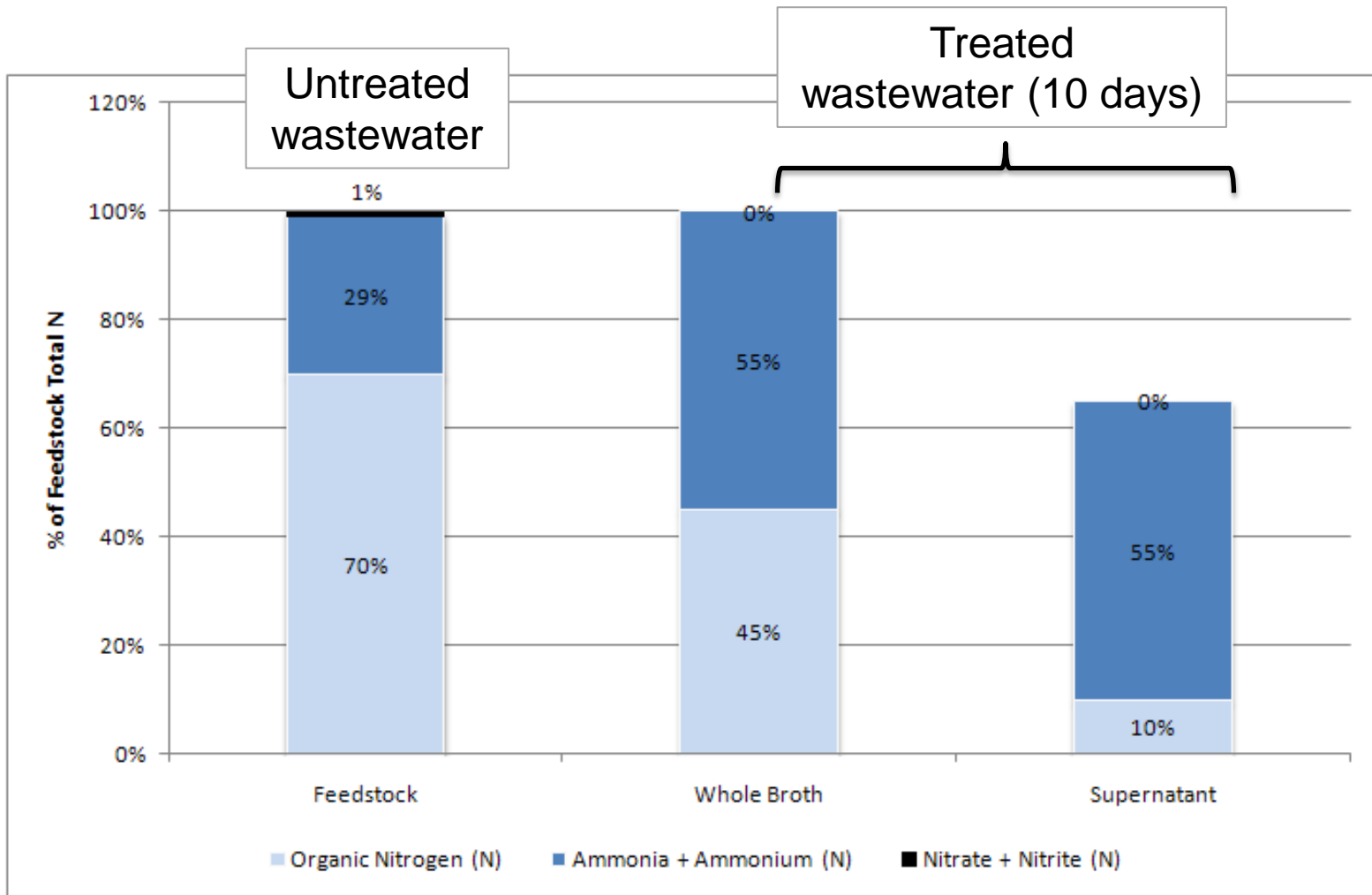
# Wastewater Treatment: COD

Results after 200 hr incubation:



# Wastewater Treatment: Nitrogen

Results after 200 hr incubation:



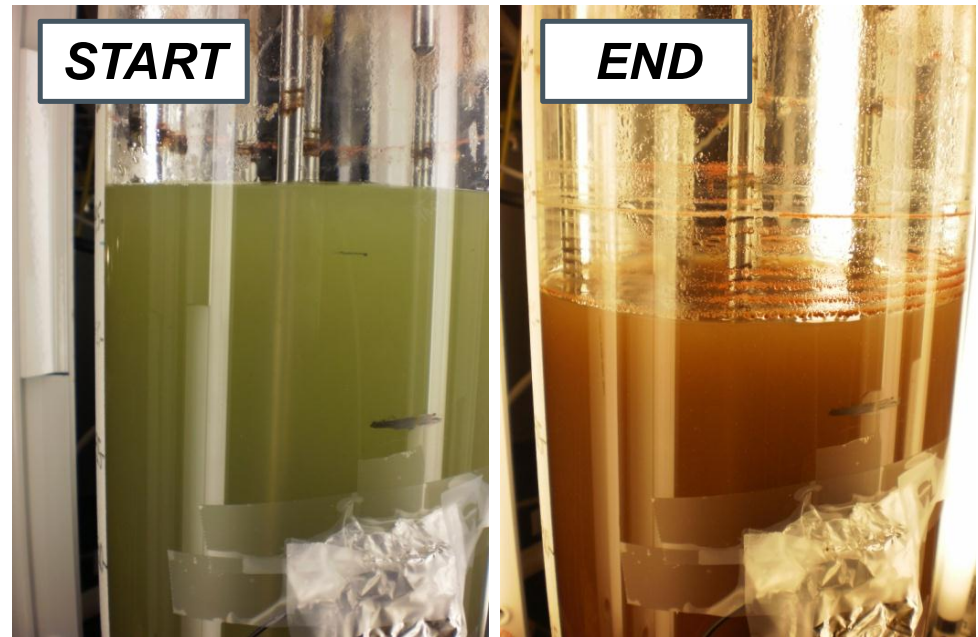
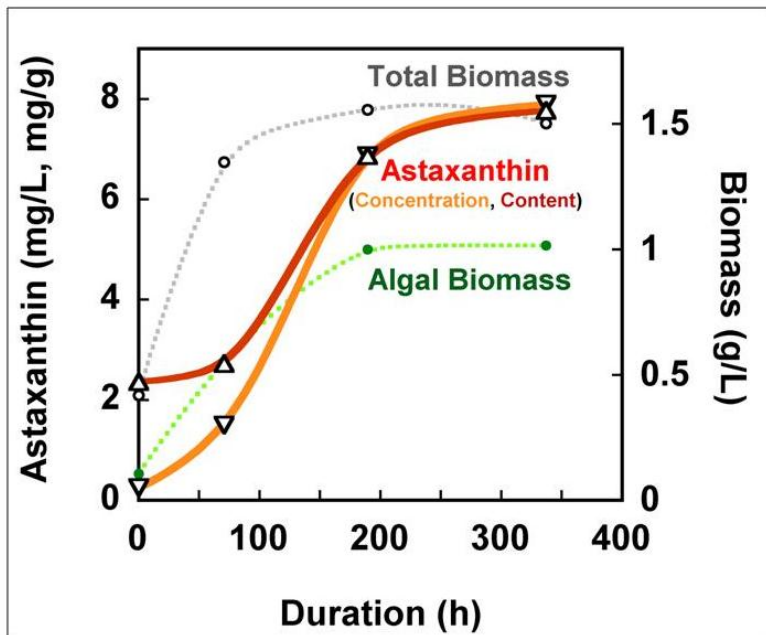
# Production of High-Value Biomass

## Results after 200 hr incubation:

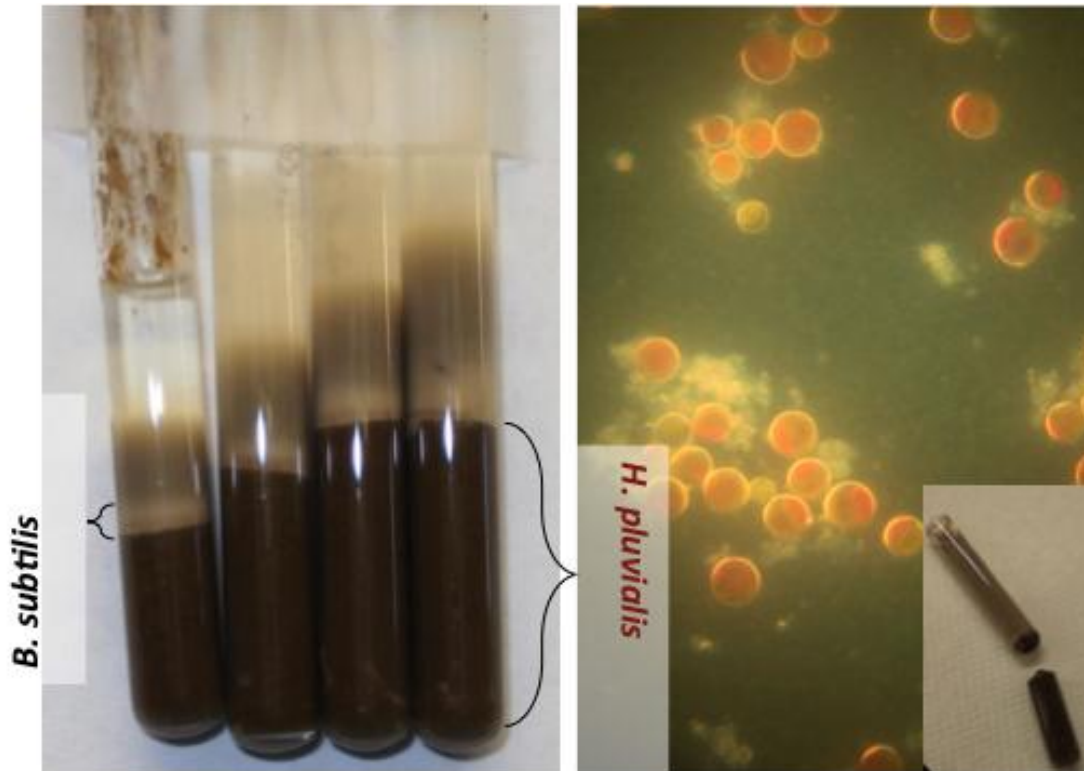
Biomass: 2.2 g/L

Algae: 1.4 g/L

**Astaxanthin: ~ 0.8%**



# Astaxanthin Accumulation



## Value proposition:

- *waste treatment (reduction in COD/BOD, N, P)*
- *high-value biomass production*
- *reduced mass-transfer, energy expenditures, as well as C emissions*

# Summary

- ❑ Phototroph-heterotroph co-cultures present an alternative option for photosynthetic production of value-added products and commodities such as biofuels.
- ❑ In comparison to axenic (pure) cultures, co-cultures display broader substrate versatility, higher productivities due to decreased mass transfer requirements, and provide increased engineering flexibility by spatially and/or temporally separating the processes of photosynthesis and photosynthate conversion
- ❑ We have successfully applied co-cultivation of heterotrophic bacteria with microalgae for wastewater treatment and production of high-value biomass. The approach opens new ways for designing highly-efficient production processes for feedstock biomass production as well as allows utilization of variety of organic agricultural, chemical, or municipal wastes.



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